

HER2-XPAT, A Novel Protease-Activated Prodrug T Cell Engager (TCE) With Potent T Cell Activation and Efficacy in Solid Tumor Models and Large Predicted Safety Margins in Non-Human Primates



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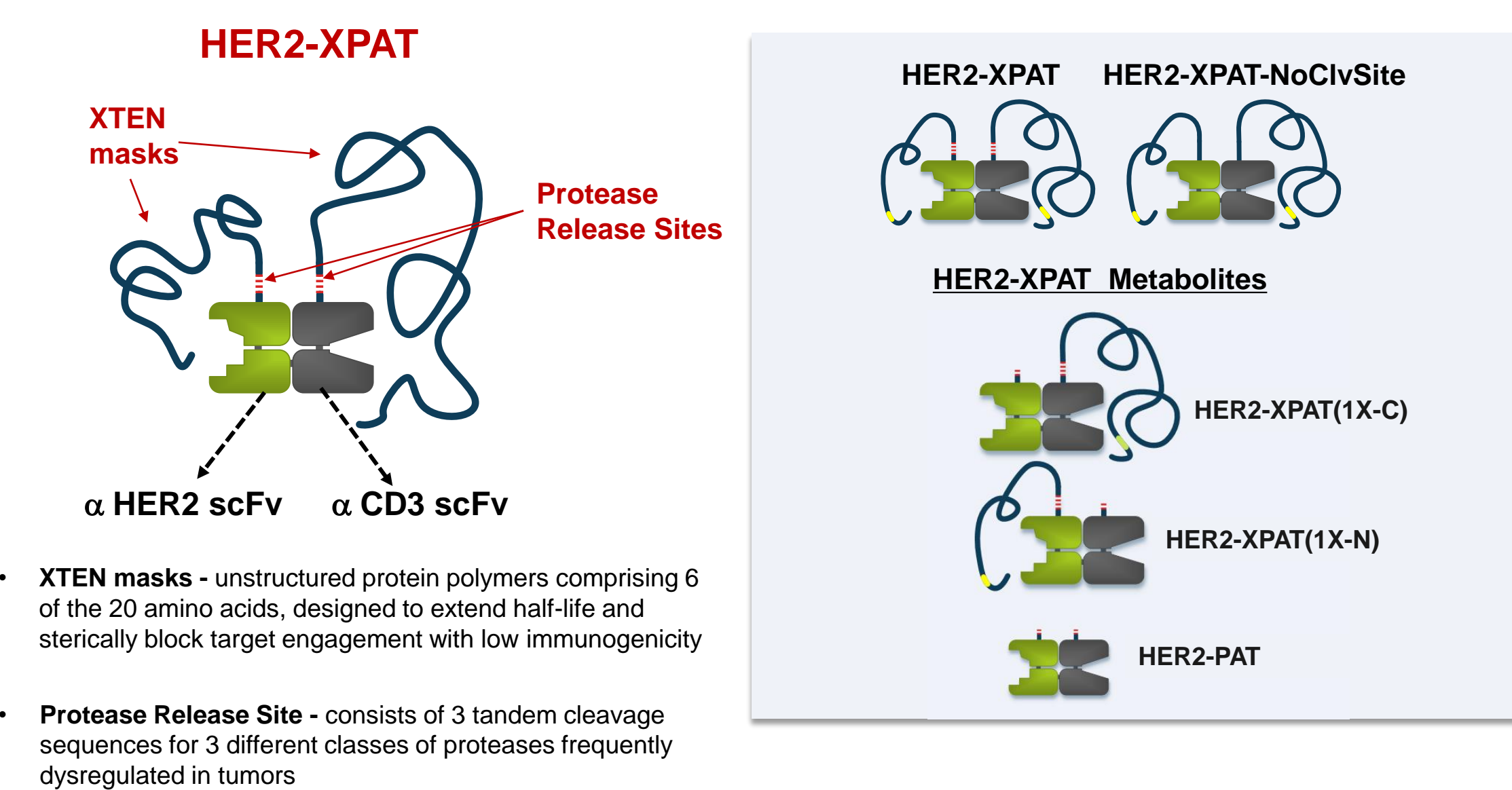
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INTRODUCTION

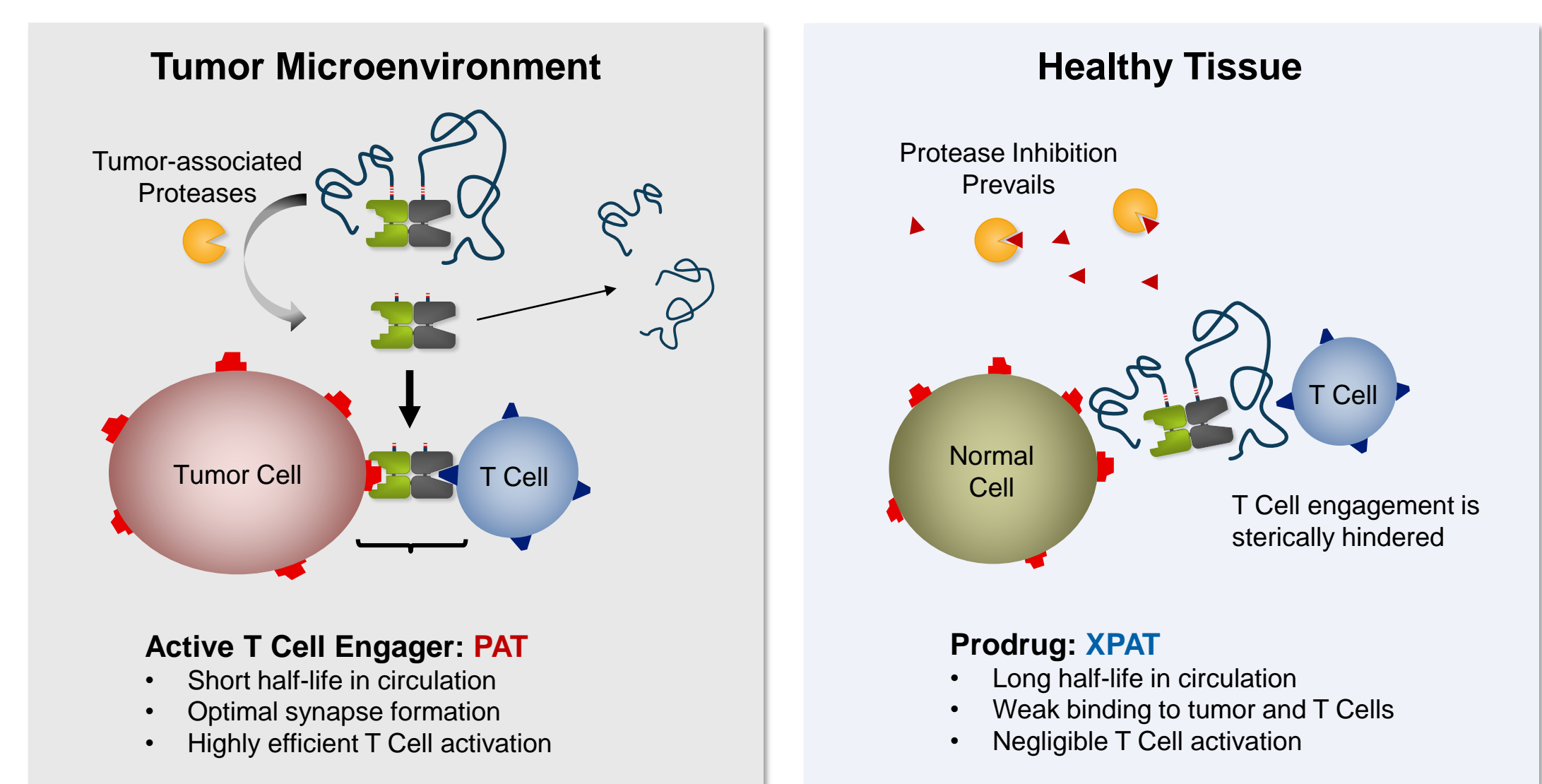
Bispecific T Cell Engagers (TCEs) have been effective at inducing remissions in hematologic cancers, but their use in solid tumors has been limited by their extreme potency and on target, off-tumor toxicities in healthy tissue. To address this challenge, Amunix has developed a conditionally-activated TCE, XPAT or XTENylated Protease-Activated bispecific T Cell Engager targeting HER2 that exploits the dysregulated protease activity present in tumors vs. healthy tissues, enabling expansion of the therapeutic index. The XPAT core consists of 2 single chain antibody fragments (scFvs) targeting CD3 and the tumor target. Two unstructured polypeptide masks (XTEN) are attached to the core that sterically reduce target engagement and extend protein half-life. Protease cleavage sites at the base of the XTEN masks enable proteolytic activation of XPAT in the tumor microenvironment, unleashing a small, highly potent TCE. In healthy tissues, where protease activity is tightly regulated, XPATs should remain predominantly inactive as intact prodrugs. In addition to localized activation, the short half-life of the unmasked PAT form should further widen the therapeutic index while providing the potency of T-cell immunity to potentially improve the eradication of solid tumors.

XPAT PLATFORM

XPATs Are XTENylated Protease-Activated T Cell Engagers

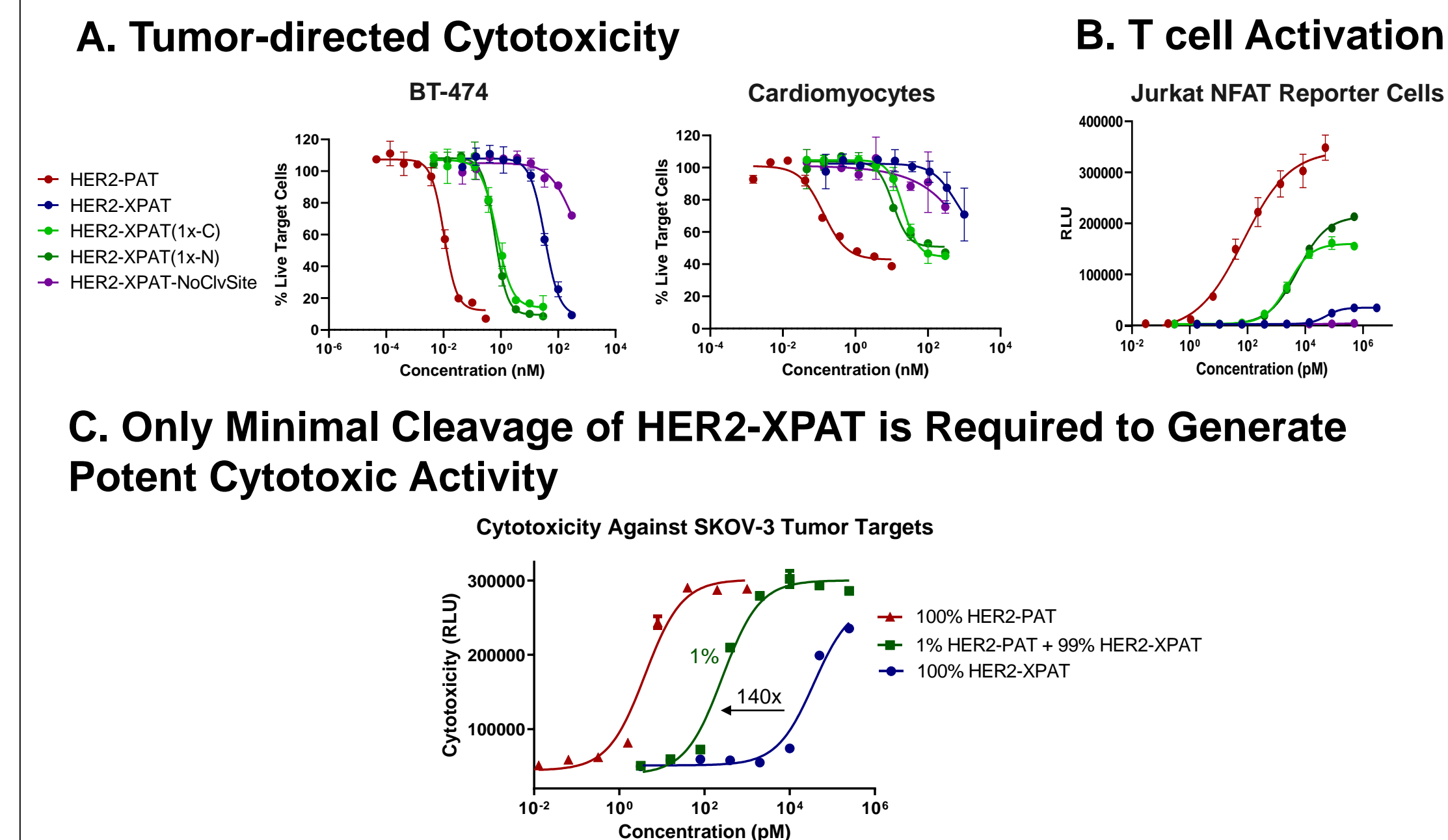


XPATs Enable Localized Tumor Killing, Limiting Toxicity Against Healthy Tissue Expressing the Target Antigen



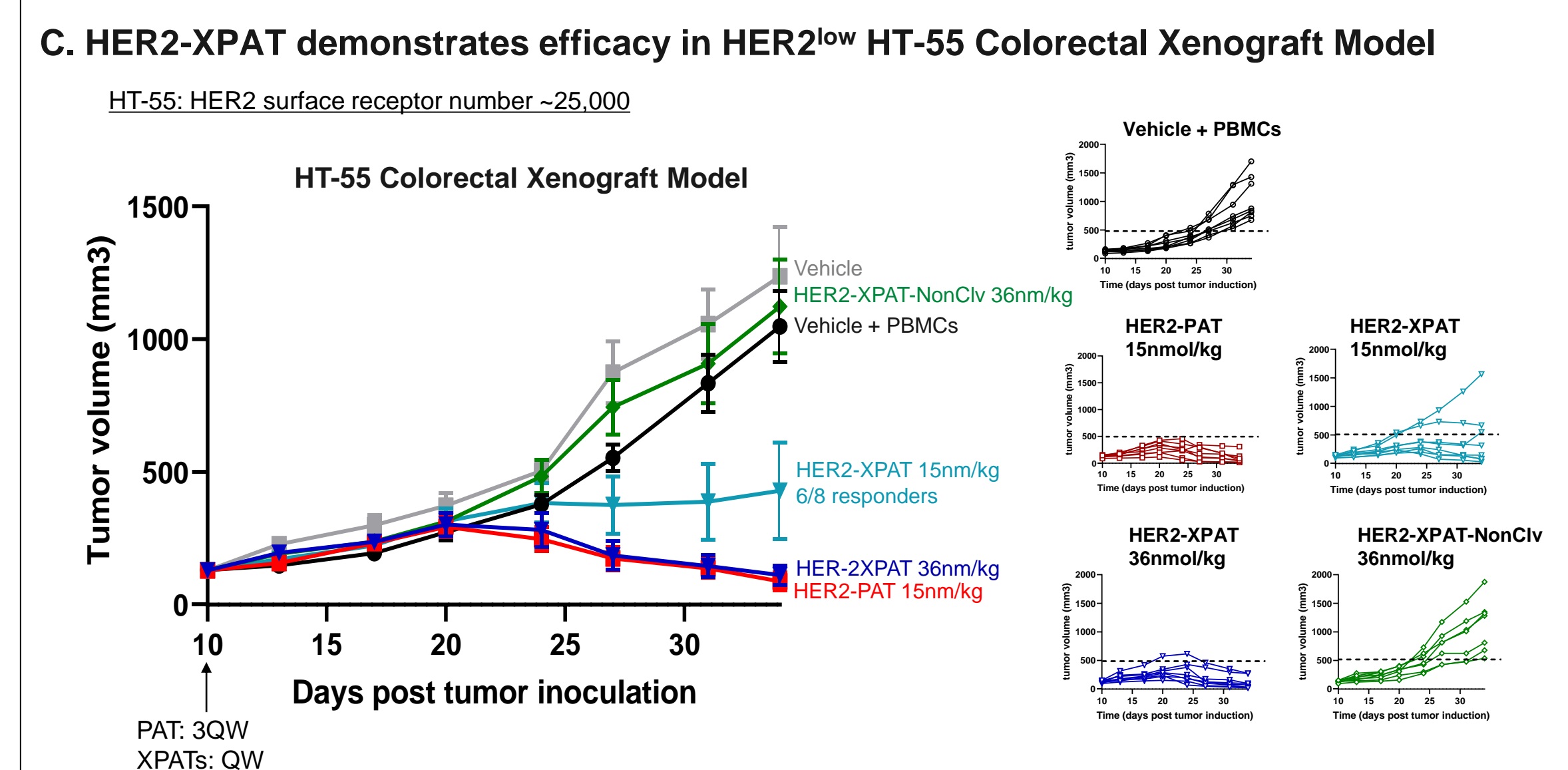
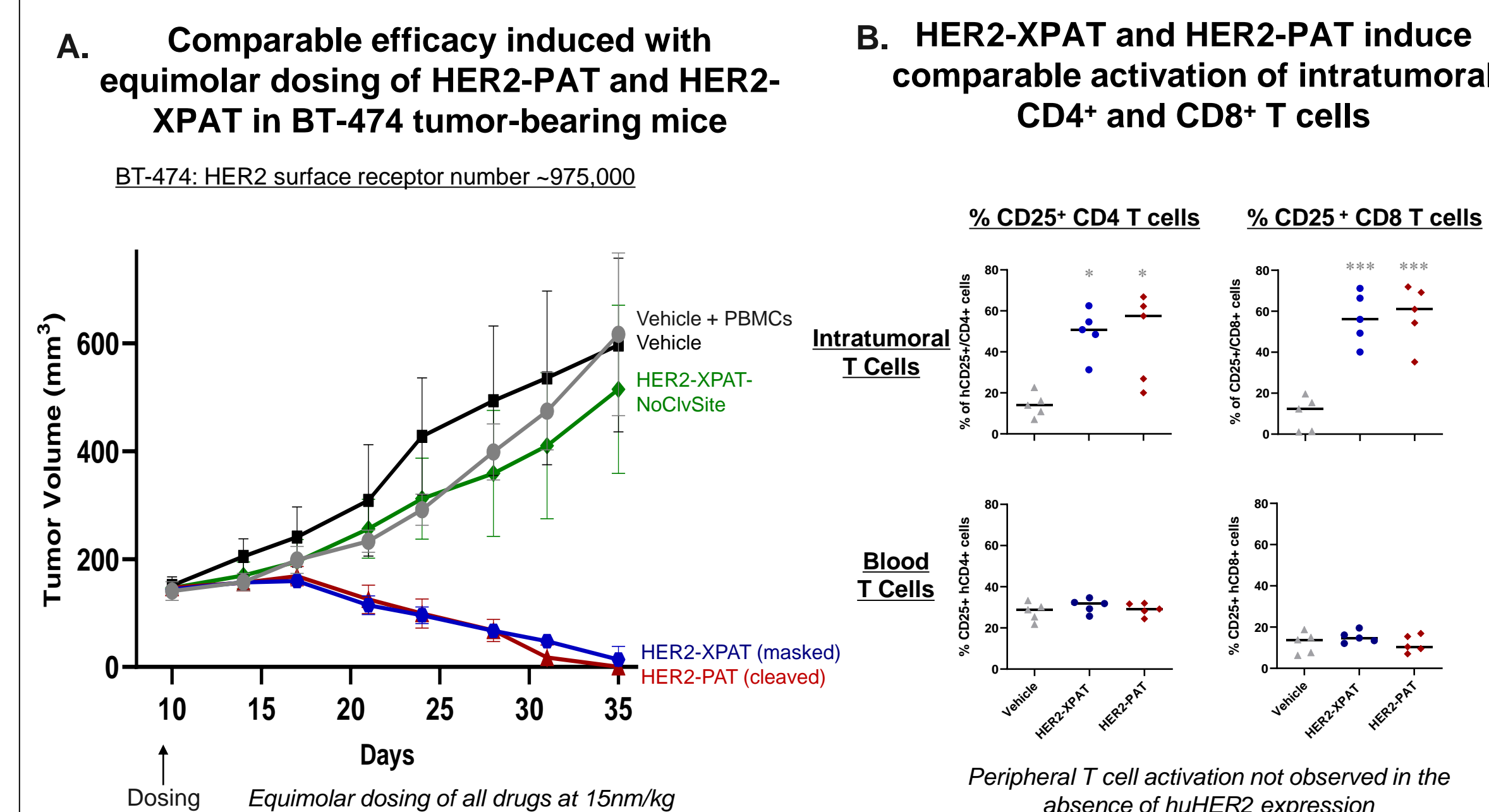
RESULTS

Figure 1. XTEN Polypeptide Masks on HER2-XPAT Significantly Reduce T Cell-Mediated Cytotoxicity and T Cell Activation *in vitro*



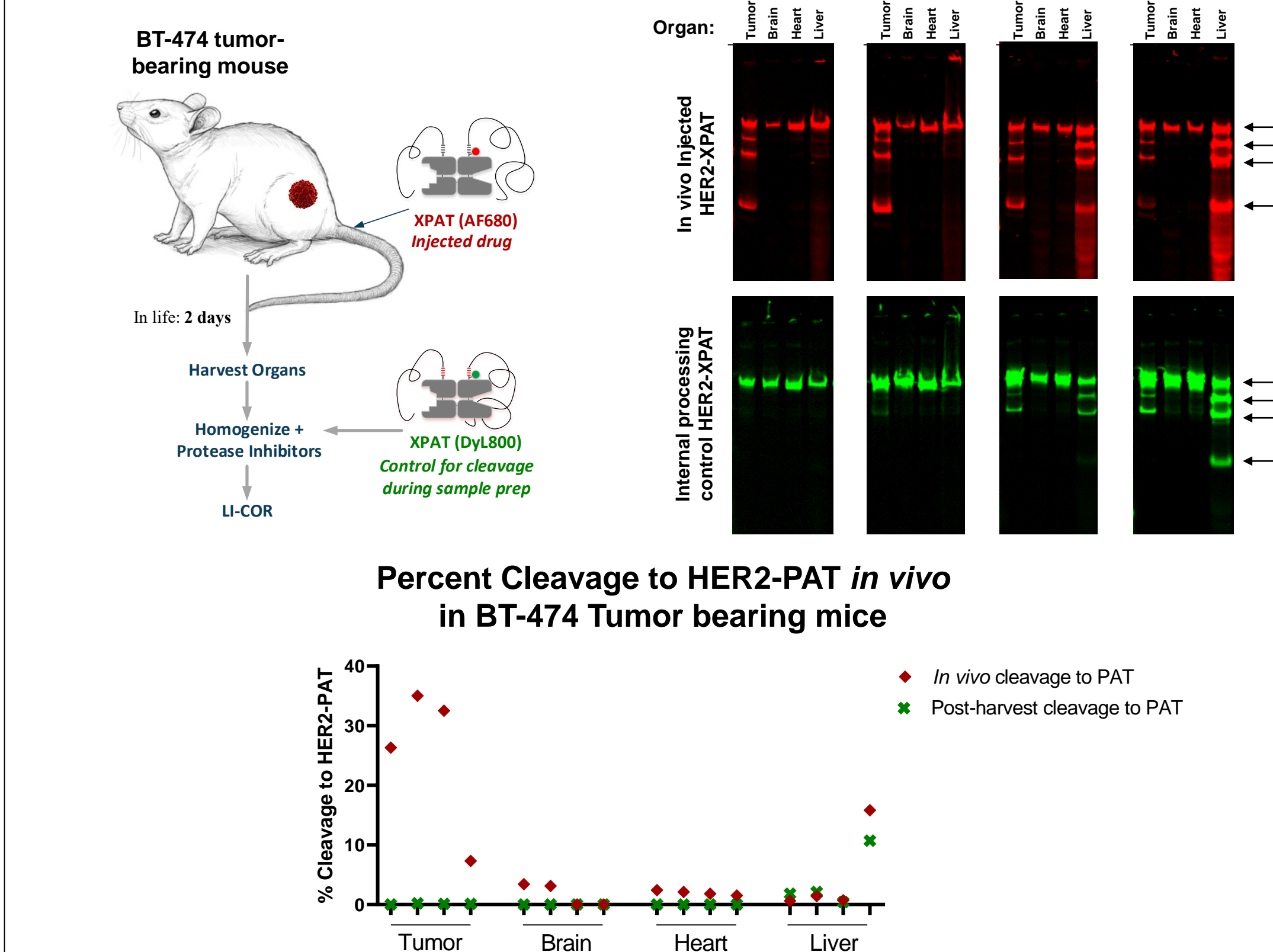
A) Cytotoxicity was quantified using Cell Titre-Glo Luminescent Cell Viability Assay following a 48 hour incubation of huPBMCs and the BT-474 tumor line or human iCell Cardiomyocytes at a 1:1 Effector:Target ratio. Co-cultures were treated with HER2-XPAT or its indicated metabolites at the concentrations shown. B) Jurkat reporter T cells were incubated with BT-474 cells and the indicated test articles at a 5:1 E:T ratio for 6 hours and NFAT-induced Luciferase activity quantified by Luminescence. C) Cytotoxicity was evaluated with SKOV3 tumor target cells in response to the indicated concentrations of HER2-XPAT and HER2-PAT and a test article composed of 1% HER2-XPAT/99% HER2-PAT.

Figure 2. HER2-XPAT Induces Robust Tumor Regressions in Mice in Both HER2^{high} and HER2^{low} Xenograft Models That Are Dependent on the Protease Release Site



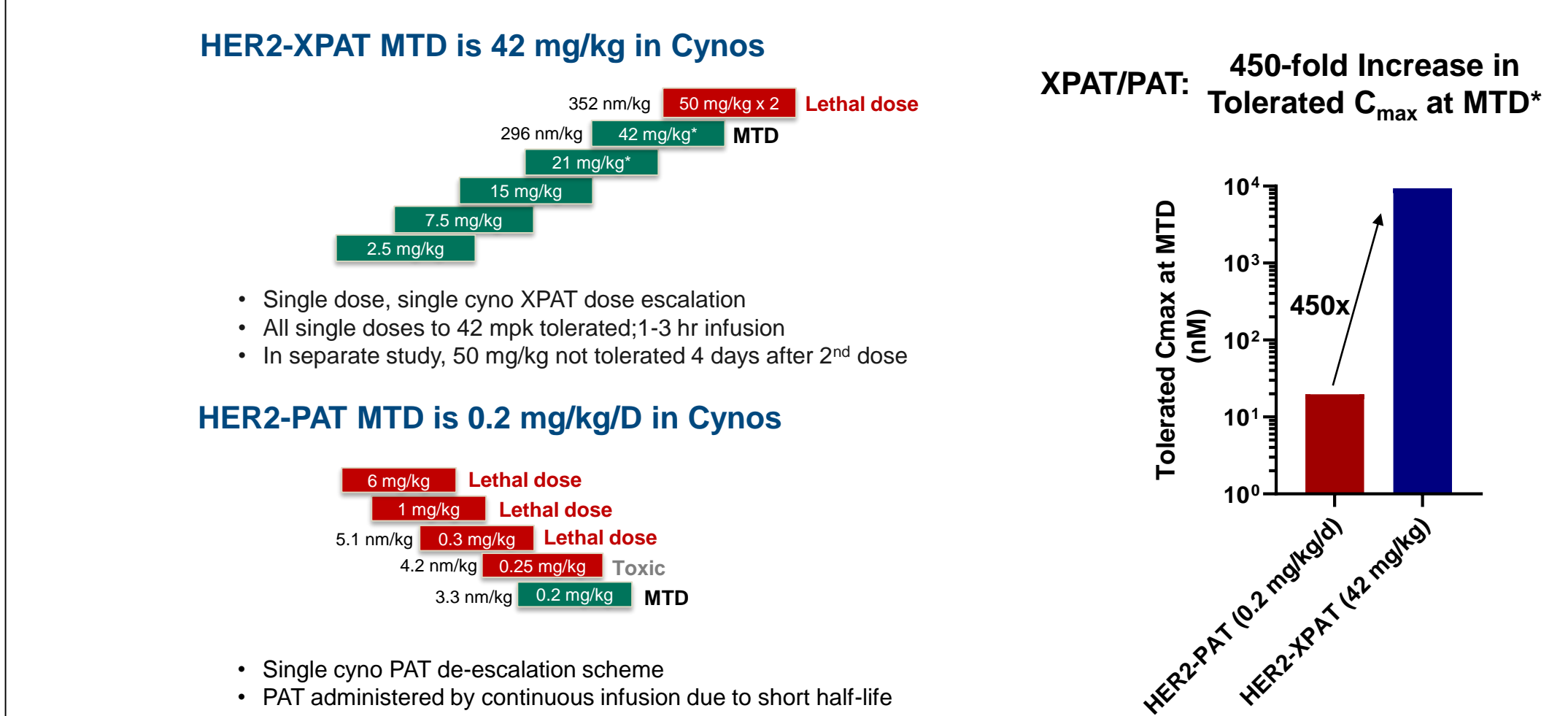
A) NOG mice were inoculated SC with 2×10^7 BT-474 tumor cells, engrafted with 1×10^7 huPBMCs on Day 8 and treated 2 days later at MTV of 147 mm^3 with the indicated test articles at equimolar doses QW for 3 weeks. Lack of tumor growth inhibition by the Non-cleavable HER2-XPAT-NoCivSite demonstrates the requirement of protease cleavage for XPAT efficacy. B) From an independent BT-474 efficacy experiment conducted as in A), the activation status of tumor infiltrating T cells was evaluated by flow cytometry on Day 18 post-TIW dosing of HER2-XPAT and HER2-PAT at 15 nmol/kg. C. NPSG mice were inoculated SC with 5×10^6 HT-55 tumor cells, engrafted with 1×10^7 huPBMCs on Day 6 and treated on D10 (at MTV of 129 mm^3) with HER2-XPAT and HER2-XPAT-NonCiv at 15 and 36 nmol/kg doses QW for 3 weeks. PAT was administered at 15nmol/kg 3QW. Lack of tumor growth inhibition by the non-cleavable XPAT format demonstrates the requirement of protease cleavage for XPAT efficacy.

Figure 3. HER2-XPAT is Preferentially Unmasked in Tumor Tissues



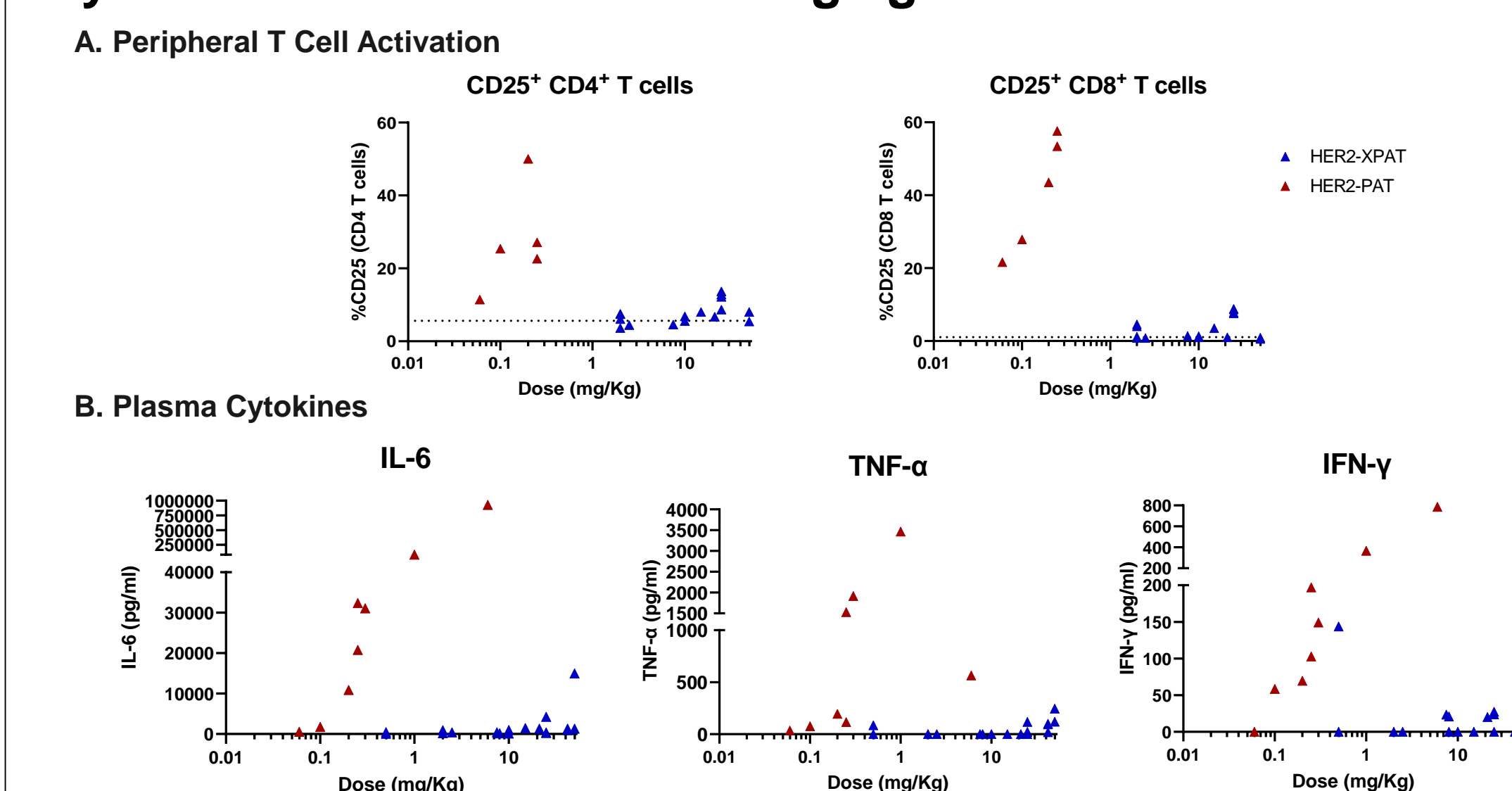
A variant of HER2-XPAT was constructed containing an additional cysteine between HER2-PAT and the C-terminal protease release site. The fluorescent dyes Alexa Fluor 680 and DyLight 800 were conjugated to this variant and the AF 680 form (red) injected into mice bearing BT-474 tumors for 2 days. The indicated organs were harvested and homogenized in the presence of protease inhibitors and the DyLight 800-labeled internal control (green). Homogenates were separated by SDS-PAGE and scanned on a LI-COR Odyssey imager. Bands corresponding to the indicated metabolites were quantified, subtracting the % cleavage obtained during processing from that obtained *in vivo*. An average of 25.2% cleavage to HER2-PAT was observed in tumor homogenates.

Figure 4. XTEN Masks Significantly Expand Safety Margin of HER2-XPAT vs. PAT in Cynomolgus Monkeys



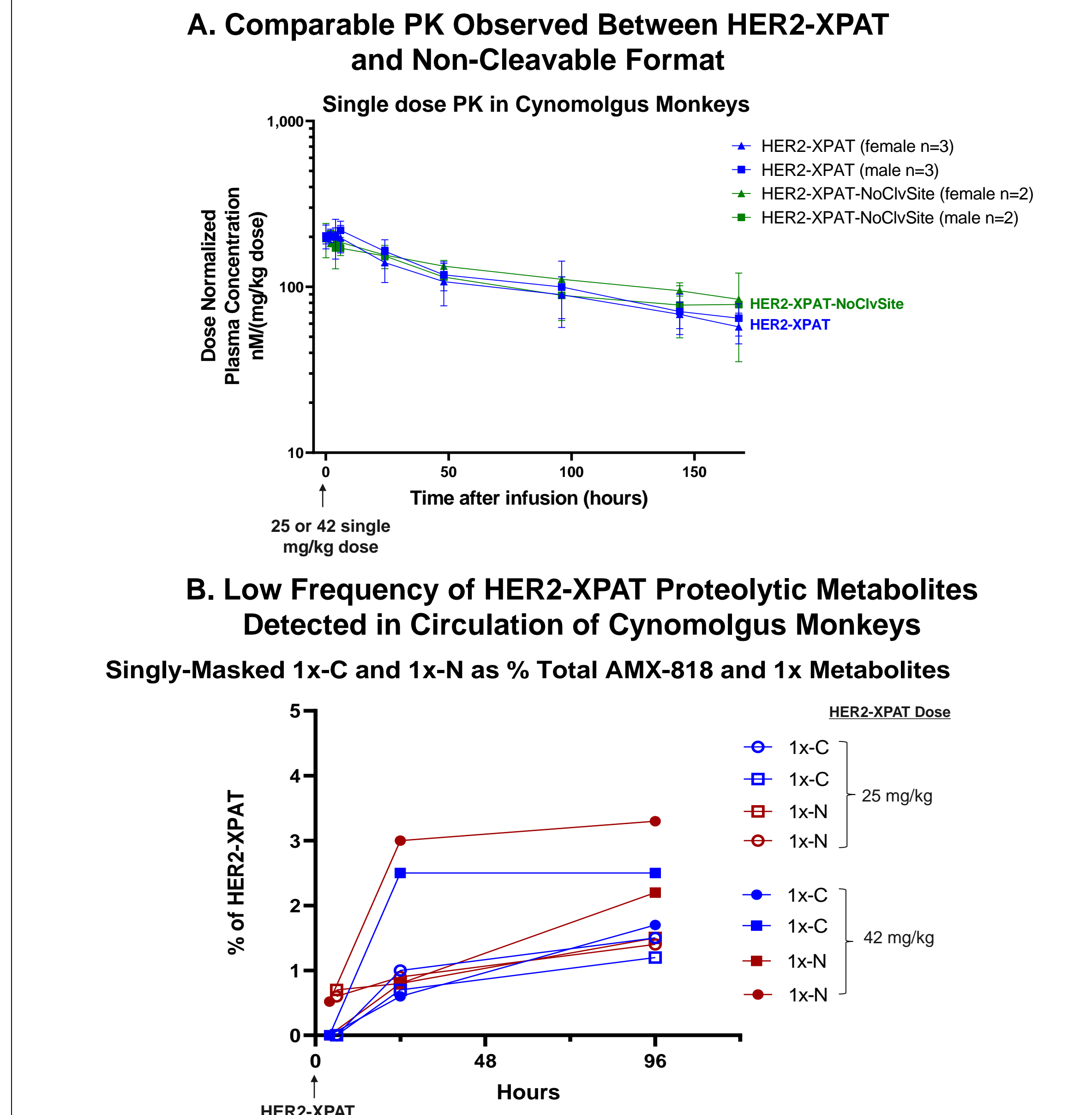
HER2-XPAT was administered IV, single dose/animal (doses 2.5-42mpk) and weekly x2 at 50mpk. *At doses 21mpk and above, a variant of HER2-XPAT with a shorter C-terminal mask was used. HER2-PAT was administered by continuous infusion due to its short half-life. Plasma concentrations of HER2-XPAT were measured by Electrochemiluminescent Immunoassay (ECLIA) using recombinant HER2 capture and an antibody directed against the XTEN masks for detection. The C_{max} values for HER2 PAT were determined by ECLIA utilizing an a-idiotypic Ab directed against the a-CD3 scFv as capture and recombinant HER2 as detection. *The 450-fold value derives from an average value for C_{max} for animals dosed with HER2-XPAT at 42mg/kg and the updated assessment for HER2-PAT MTD at 0.2 mg/kg/day.

Figure 5. Masks on HER2-XPAT Provide Significant Protection from Peripheral T Cell Activation and Cytokine Release Syndrome in NHP Even at 50mg/kg



A) Peripheral T cell activation (%CD25⁺) was evaluated by flow cytometry 24 hours post-HER2-XPAT treatment. B) Cytokine analysis was performed with a Luminex® suspension array system on plasma samples. Data presented are maximal values measured between 6-24 hours at each evaluated dose.

Figure 7. HER2-XPAT Protease Release Site Remains Largely Stable in Circulation of Cynomolgus Monkeys Even at High Doses, Consistent With its Strong Safety Profile



A) HER2-XPAT or its non-cleavable counterpart, HER2-XPAT-NoCivSite were administered as a single IV dose of 25 or 42 mg/kg. Dose normalized plasma drug concentrations were measured by ECLIA using recombinant HER2 as capture and an antibody directed against the XTEN mask for detection. *ECLIA = Electrochemiluminescent Immunoassay. B) Concentrations of singly-cleaved HER2-XPAT(1X-C) and HER2-XPAT(1X-N) were measured in plasma from cynomolgus monkeys administered HER2-XPAT. Quantitative Western was performed using an antibody recognizing the anti-HER2 scFv with standards prepared with recombinant versions of the singly-cleaved molecules. Results are expressed as a percentage of the total XTENylated species as measured by ECLIA as described above.

SUMMARY/CONCLUSIONS

- *In vitro*, proteolytically-unmasked HER2-PAT demonstrates potent cytotoxicity against tumor lines with EC50s in the single-digit pM range. Double XTEN masking reduces target-directed T cell cytotoxicity and T cell activation by up to 4 orders of magnitude, while singly-masked XPATs show intermediate activity relative to unmasked HER2-PAT. Only minimal cleavage of XPAT is required to generate potent cytotoxicity
- In the established HER2^{high} BT-474 and HER2^{low} HT-55 xenograft models, HER2-XPAT induced protease-dependent tumor regressions comparable to the unmasked (active) T cell engager while remaining stable in circulation. *In vivo*, preferential cleavage of HER2-XPAT was demonstrated in tumors relative to healthy organs (average % HER2-PAT was 25.2% in tumors and 1.6% in combined other organs)
- In cynomolgus monkeys, HER2-XPAT demonstrated a high safety margin, supported by its protease stability in circulation and a maximum tolerated exposure that was ~450-fold higher than that of its active form (PAT). No CRS or systemic T cell activation was observed even at 50 mg/kg, supportive of minimal CRS risk for XPATs vs standard TCEs. Only 1-3% of singly-cleaved XPAT metabolites were detected in plasma from NHP administered high doses of HER2-XPAT (25 & 42mg/kg)
- XPATs represent a novel strategy to improve the toxicity profile of T cell engagers while maintaining their potency against solid tumors, thus enabling a significant increase in the therapeutic index and expansion of target landscape for this potent modality